# Package: RTIGER (via r-universe)

August 26, 2024

Type Package Title HMM-Based Model for Genotyping and Cross-Over Identification Version 2.1.0 **Description** Our method integrates information from all sequenced samples, thus avoiding loss of alleles due to low coverage. Moreover, it increases the statistical power to uncover sequencing or alignment errors <doi:10.1093/plphys/kiad191>. **Depends** R (>= 3.6), GenomicRanges, GenomeInfoDb License GPL (>= 2) **Encoding** UTF-8 LazyData true LazyDataCompression gzip Imports methods, e1071, extraDistr, reshape2, ggplot2, TailRank, JuliaCall, IRanges, qpdf, grDevices, graphics, stats, utils RoxygenNote 7.2.3 VignetteBuilder knitr Suggests knitr, rmarkdown, markdown, Gviz, rtracklayer biocViews GenomeAnnotation, HiddenMarkovModel, Sequencing Repository https://rfael0cm.r-universe.dev RemoteUrl https://github.com/rfael0cm/rtiger RemoteRef HEAD RemoteSha c187e841133cf0ca1cb3da026f108f86aca4f5e1

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ATseqlengths

The autosome chromosome lengths for Arabidopsis Thaliana.

#### Description

The autosome chromosome lengths for Arabidopsis Thaliana.

#### Author(s)

Rafael Campos-Martin

calcCOnumber Obtain number of Cross-Over events per sample and chromosome.

#### Description

Obtain number of Cross-Over events per sample and chromosome.

# Usage

```
calcCOnumber(object)
```

#### Arguments

object a RViterbi object.

#### Value

Matrix m x n. M number of samples and N chromosomes.

#' @return a matrix with n chromosomes and m samples (n x m) and the number of CO events.

#### Examples

```
data("fittedExample")
co.num = calcCOnumber(myDat)
```

dev

#### Description

Function to developers. It runs one EM step

#### Usage

```
dev(psi, rigidity = NULL, nstates = 3, transition = NULL, start = NULL)
```

#### Arguments

psi	list of psi probabilities.
rigidity	Rigidity value.
nstates	Number of states.
transition	transition matrix
start	initial probabilities

# Value

List with updates probabilites

fit

Call Julia code to fit the values

# Description

Call Julia code to fit the values

#### Usage

```
fit(rtigerobj, max.iter , eps,
trace, all = TRUE, random = FALSE,
specific = FALSE, nsamples = 20,
post.processing = TRUE)
```

#### Arguments

rtigerobj	an RTIGER object.				
max.iter	maximum number of iterations to acomplish by the EM.				
eps	differnece threshold to halt the EM.				
trace	logical value whether to trace the changes in the parameters along the iterations.				
all	logical value whether to use all data to fit the model.				
random	if all FALSE use random samples.				
specific	if all FALSE use specific samples.				
nsamples	if random TRUE, how many samples to use.				
post.processing					
	logical value, whether to run post.processing process.				

#### Value

**RTIGER** object

# Examples

```
## Not run:
data("fittedExample")
sourceJulia()
myfit = fit(myDat, max.iter = 2, eps=0.01,
            trace = TRUE, all = TRUE,
            random = FALSE, specific = FALSE,
            nsamples = 20, post.processing = TRUE)
```

## End(Not run)

generateObject Load data

#### Description

Load data

#### Usage

```
generateObject(experimentDesign = NULL,nstates = 3, rigidity=NULL,
seqlengths = NULL, verbose = TRUE)
```

#### myDat

#### Arguments

experimentDesign				
	a data Frame that contains minimum a column with the files direction (name of the column files) and another with a shorter name to be used inside the function.			
nstates	the number of states to be fitted in the model. A standard setting would use 3 states (Homozygous1, Heterozygous, and Homozygous2).			
rigidity	an integer number specifying the rigidity parameter to be used.			
seqlengths	a named vector with the chromosome lenghts of the organism that the user is working with.			
verbose	logical value. Whether to print info messages.			

#### Value

**RTIGER** object

#### Examples

myDat

A fitted example using three own samples of Arabidopsis. More information in publication:

#### Description

A fitted example using three own samples of Arabidopsis. More information in publication:

#### Author(s)

Rafael Campos-Martin

optimize\_R

# Description

Find the otimum R value for a given data set

#### Usage

```
optimize_R(object,
max_rigidity = 2^9, average_coverage = NULL, crossovers_per_megabase = NULL,
save_it = FALSE, savedir = NULL)
```

# Arguments

object	an RTIGER object
<pre>max_rigidity</pre>	R values will be explored up the value given in this parameter. Default = $2^{9}$
average_covera	ige
	For conservative results set it to the lowest average coverage of a sample in your experiment, or evne to the lowest average coverage in a (sufficiently large) region in one of your samples. The lower the value, the more conservative (higher) our estimates of the false positive segments rates. If it is not provided it will be computed as the average of all data points.
crossovers_per	megabase
	For conservative results set it to the highest ratio of a sample in your experiment.
	The higher the value, the more conservative (higher) our estimates of the false
	positive segments rates. If it is not provided it will be computed as the average of all samples.
save_it	logical values if the results should be saved. Plots might be complicated to inter-
	pret. We suggest to read the manuscript to understand them (https://doi.org/10.1093/plphys/kiad191)
savedir	if results are saved, in which directory.

#### Value

A value with the optimum rigidity for the data set.

# Examples

```
data("fittedExample")
bestR = optimize_R(myDat)
```

plotCOs

#### Description

Obtain number of Cross-Over events per sample and chromosome.

#### Usage

```
plotCOs(object, file = NULL)
```

#### Arguments

object	a RViterbi object.
file	file where to save the plot for CO numbers

#### Value

a plot

#### Examples

```
data("fittedExample")
co.num = calcCOnumber(myDat)
```

RTIGER

Load, Fit, and plot

#### Description

Load, Fit, and plot

#### Usage

```
RTIGER(expDesign, rigidity=NULL, outputdir=NULL, nstates = 3,
seqlengths = NULL, eps=0.01, max.iter=50, autotune = FALSE,
max_rigidity = 2^9, average_coverage = NULL,
crossovers_per_megabase = NULL, trace = FALSE,
tiles = 4e5, all = TRUE, random = FALSE, specific = FALSE,
nsamples = 20, post.processing = TRUE, save.results = TRUE, verbose = TRUE)
```

# Arguments

expDesign	a data Frame that contains minimum a column with the files direction (name of the column files) and another with a shorter name to be used inside the function.					
rigidity	an integer number specifying the rigidity parameter to be used.					
outputdir	a character string that specifies the directory in which to save the results form the function.					
nstates	the number of states to be fitted in the model. A standard setting would use 3 states (Homozygous1, Heterozygous, and Homozygous2).					
seqlengths	a named vector with the chromosome lenghts of the organism that the user is working with.					
eps	the threshold of the difference between the parameters value between the previ- ous and actualy iteration to stope de EM algorithm.					
max.iter	maximum number of iterations of the EM algorithm before to stop in case that eps has not been achieved.					
autotune	Logical value if the R-value should be tuned by our algorithm. This will take longer as it needs a first training with the rigidity value provided by the user and then the optimization step is carried. Finally, a training using the optimum R will be performed and results for the optimum R will be returned.					
<pre>max_rigidity</pre>	If autotune true, R values will be explored up the value given in this parameter. Default = $2^{9}$					
average_coverag						
	If autotune true, for conservative results set it to the lowest average coverage of a sample in your experiment, or evne to the lowest average coverage in a (sufficiently large) region in one of your samples. The lower the value, the more conservative (higher) our estimates of the false positive segments rates. If it is not provided it will be computed as the average of all data points.					
crossovers_per_						
	If autotune true, for conservative results set it to the highest ratio of a sample in your experiment. The higher the value, the more conservative (higher) our estimates of the false positive segments rates. If it is not provided it will be computed as the average of all samples.					
trace	logical value. Whether or not to keep track of the parameters for the HMM along the iterations. Deafault FALSE					
tiles	length of the tiles by which the genome will be segmented in order to compute the ratio of COs in the complete dataset.					
all	logical value. Whether to use the complete data set to fit the rHMM. default TRUE.					
random	Logical value. Choose randomly a subset of the complete dataset to fit the rHMM. Default FALSE					
specific	Logical value to specify which samples to take.					
nsamples	if random TRUE, how many samples should be taken randomly.					
post.processing						
	Logical value. Whether to run an extra step that fine maps the segment borthers. Default TRUE					

#### **RTIGER-class**

save.results	Logical value, whether to generate and save the plots and igv files.
verbose	Logical, whether to print info to console.

#### Value

Matrix m x n. M number of samples and N chromosomes. RTIGER object

#### Examples

## End(Not run)

RTIGER-class

This class is a generic container for RTIGER analysis

#### Description

This class is a generic container for RTIGER analysis

#### Slots

matobs Nested lists. the first level is a list of samples. For each sample there are 5 matrices that contains the allele counts for each position.

params a list with the parameters after training.

info List with phenotipic data of the samples.

Viterbi List of chromosomes with the viterbi path per sample.

Probabilities Computed probabilites for the EM algorithm.

num.iter Number of iterations needed to stop the EM algorithm.

setupJulia

#### Description

Installs the needed packages in JULIA to run the EM algorithm for rHMM.

#### Usage

setupJulia(JULIA\_HOME = NULL)

#### Arguments

JULIA\_HOME the file folder which contains julia binary, if not set, JuliaCall will look at the global option JULIA\_HOME, if the global option is not set, JuliaCall will then look at the environmental variable JULIA\_HOME, if still not found, JuliaCall will try to use the julia in path.

#### Value

empty

sourceJulia	Function needed before using RTIGER() function. It loads the scripts
	in Julia that fit the rHMM.

# Description

Function needed before using RTIGER() function. It loads the scripts in Julia that fit the rHMM.

#### Usage

sourceJulia()

#### Value

empty

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